

X-ray Crystallography of HLA Single Chain Trimers for Therapeutics Targeting HPV-induced Cancers

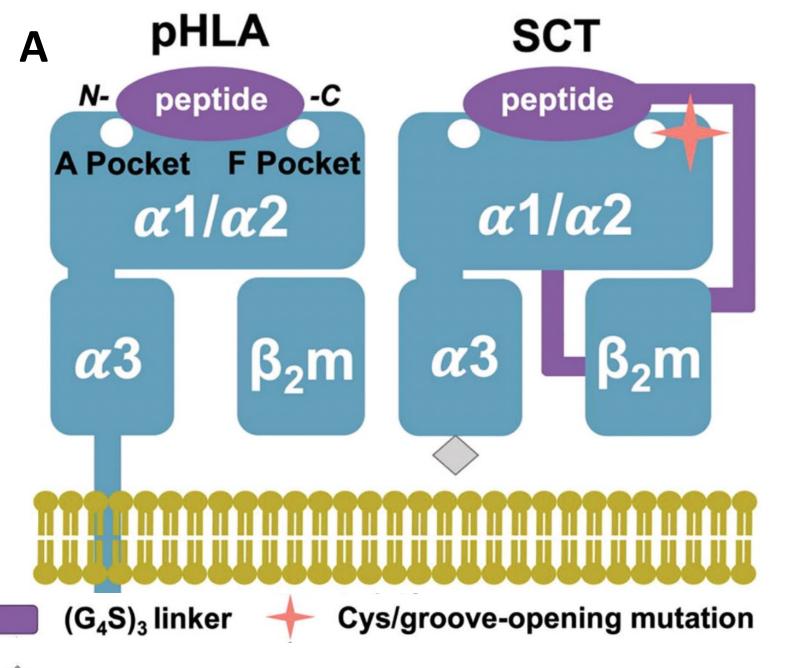
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Background and Motivation

- Human major histocompatibility complex (MHC) class I proteins or human leukocyte antigens (HLA-I) present peptide fragments from endogenous proteins on the cell surface for recognition by T cell receptors (TCRs) and NKG2x / CD49 or KIR natural killer cell receptors.¹
- Single chain trimers (SCTs) couple all three components of peptide/HLA-I complex (pHLA) (the integral-membrane heavy- or α -chain, the invariant light β_2 -microglobin chain, and the peptide fragment) into a single polypeptide, allowing recombinant expression via secretion from eukaryotic cells.¹
- Immunotherapies can be designed to bind to pHLA complexes presented on the surface of HPV induced tumor cells based on X-ray crystallographic structures of SCTs.



His₆ tag

HPV peptide HLA class 1 allele sequence HLA-A*11:01 SVYGTTLEQQYNK HLA-A*03:01 SVYGTTLEQQYNK HLA-A*29:02 SVYGTTLEQQY HLA-A2 **VLDFAPPGA**

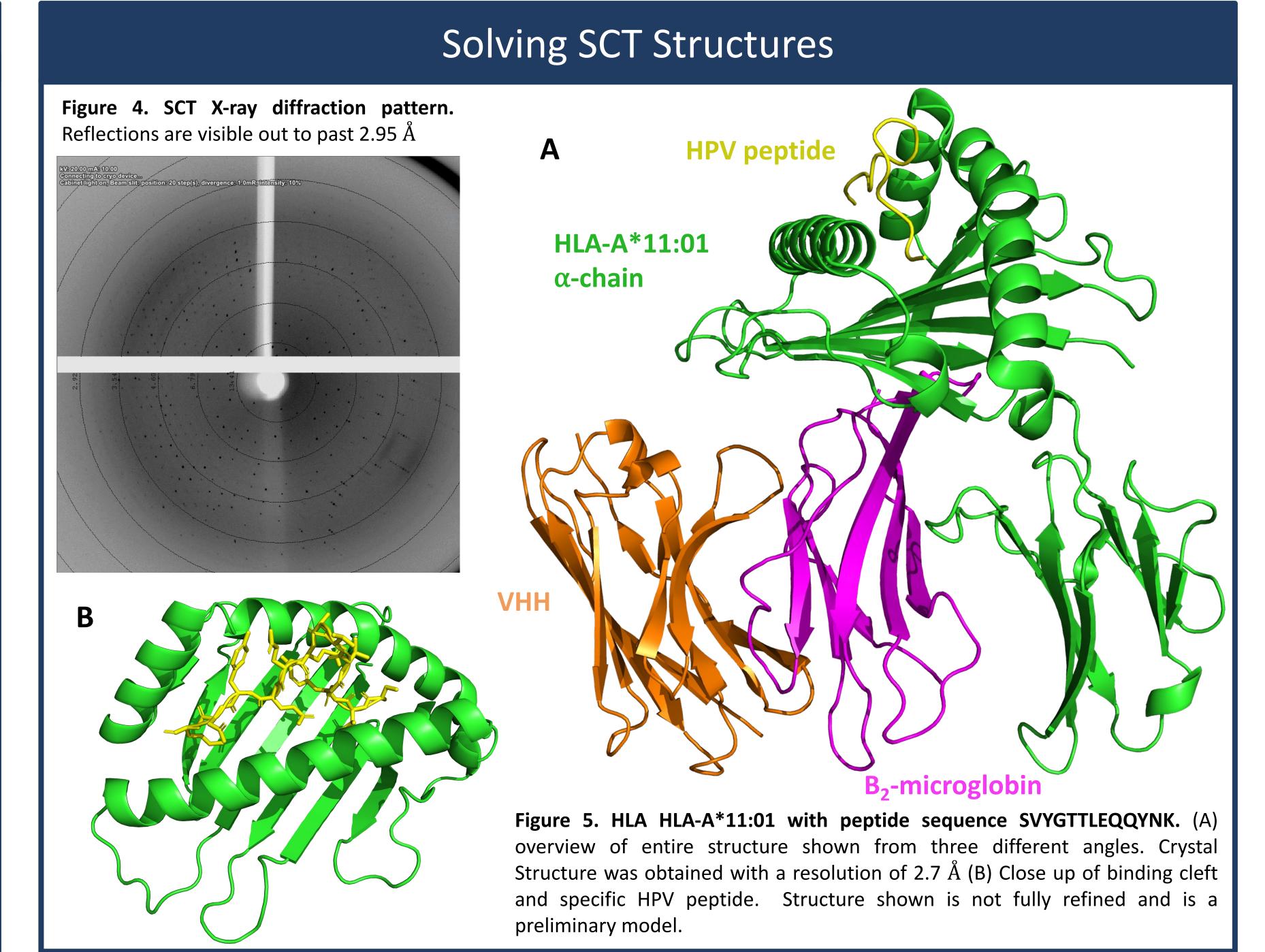
Figure 1. SCT design and structure. (A) Native pHLA Structure and SCT structure.¹ (B) Table containing the HLA alleles and peptides of the SCTs we hoped to produce and crystallize.

Crystal Screening Buffers Protein drop: 96 well screens Precipitants Addatives MES (pH=6) 96 well additive screen PEG 2k 1 μL protein @ 2-4 PEG 2k MME HEPES (pH=7) 30% v/v Dimethyl sulfoxide AmSO4 mg/mL pHClear PEG 3350 0.1 M Urea TRIS (pH=8) • 1 μL well solution PEGs I PEG 6k TRIS (pH=8.5) 30% w/v D-Sorbitol PEGs II 30% w/v D-Sorbitol Well solution (1 mL) 30% v/v Ethylene glycol 96 well screening plate 30% w/v 1,6-Hexanediol 30% v/v Ethanol 1 2 3 4 5 6 7 8 9 10 11 30% w/v Sucrose 24 well hanging Scale up for bigger crystals **Protein drop:** • 100 nL protein @ 6-8 mg/mL 100 nL well solution Well solution (80 μL): 12-28% w/v precipitant • 0.1-0.2M buffer or salt Additives

Figure 3. SCT crystallization. (A) Crystallization well condition screenings. (B) Diagram showing 96 well plate setup. At this stage the well volume is 80 μL and the drop is a 200 nL sitting drop. (C) 24 well plate set up. Here the well volume is 1 mL and the drop is a 2 μL hanging drop (D.1) Thick singular crystal. (D.2) Sheet crystals. (D.3) needle crystals.

SCT Production and Purification (1) Gibson Transduction into Lentiviral assembly Transfection of (4) Protein Purification **HEK293T cells Size Exclusion Chromatography Absorption** 158 kDa **↓ 44 kDa ↓ 88.77 mL** 73,31 mL

Figure 2. SCT expression and purification. (A) Process flow to produce SCTs for crystallization. (B) Protein PAGE gel indicating successful HisTag purification of two SCTs. Columns 1 and 4 are the protein elution. Columns 2 and 5 are the column flow through. Columns 3 and 6 are from the left-over nickel resin. (C) Size exclusion chromatography (SEC) UV absorbance [mAU] as a function of mL of PNE buffer passed through the column. Dotted lines are measured standards indicating that protein peak falls within the expected range for a ≈50 kDa protein.



Significance and Future Directions

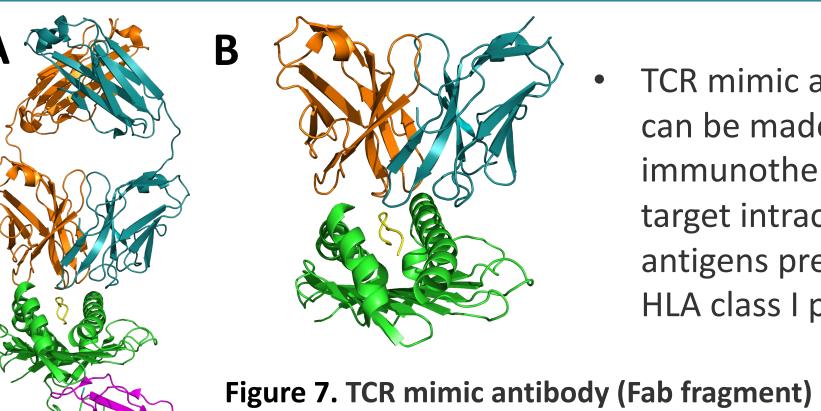
CAR/Adaptive T-cell Therapies



- Structures can inform the design of Tcell based therapeutics that target specific HLA class I complexes
- SCTs used to indicate which TCRs will bind to the target cell
- SCTs inform mutation of TCRs to create stronger binders

Figure 6. MHC class I TCR complex (4MS8).² Only the binding cleft (green) and the peptide (blue) of the MHC complex is shown. The T-cell receptor consists of two chains, the α -chain (purple) and the β -chain (yellow).

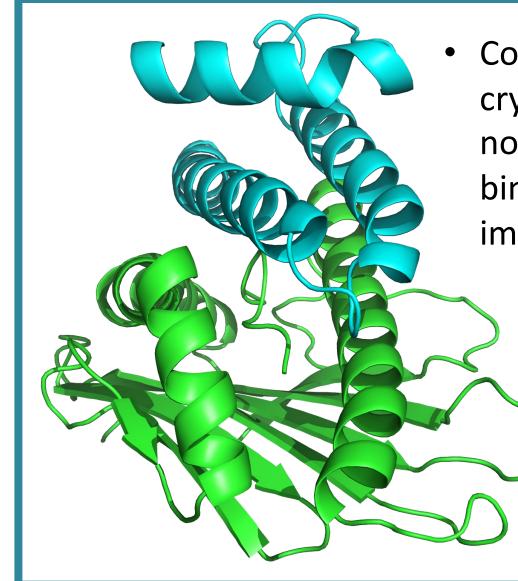
Antibody Therapies



TCR mimic antibodies can be made as immunotherapies that target intracellular antigens presented by HLA class I proteins

In complex with AFP/HLA-A*02 (7RE7).3 (A) Overview of structure. (B) Close up view of the HLA peptide cleft and binding region of the Fab

De Novo Computational Modeling



Collaborator Aaron Ring uses x-ray crystallographic structures for deimmunotherapies.

> Figure 8. Aaron Ring's model of a binder to HLA-A2 with an HPV peptide from E7 protein. Shown in green is the binding cleft and peptide of the HLA complex. Shown in blue is the designed binder.

References

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Acknowledgments

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