# Cryo-EM reveals a *de novo* minibinder induces a conformational change in α5β1 integrin

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#### BACKGROUND

#### What is integrin?

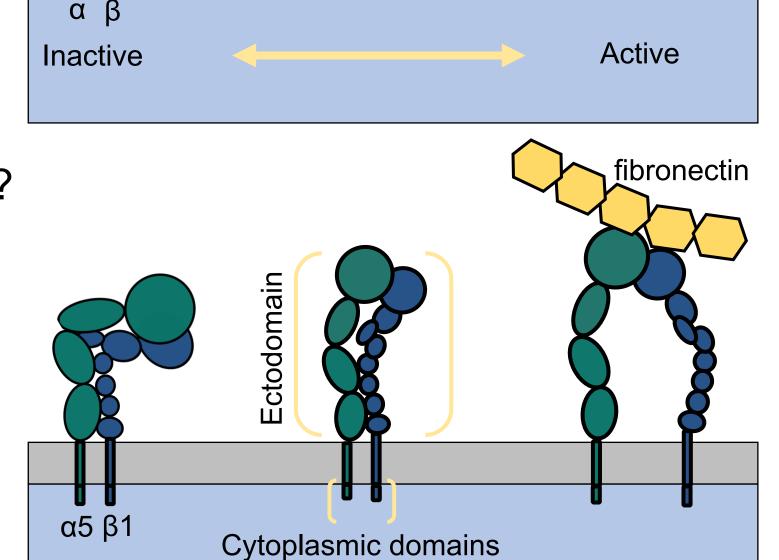
- Membrane proteins expressed on the surface of all nucleated cells<sup>1</sup>
- ❖ Go through drastic conformational changes associated with function<sup>1</sup>
- ❖ Different cations, like Mn<sup>2+</sup> can activate/induce the extended open state<sup>1</sup>

## What is $\alpha 5\beta 1$ integrin?

- ❖ Fibronectin receptor² Does not follow typical integrin conformational
- Expressed on the surface of epithelial

changes<sup>2</sup>

Main functions are in cell adhesion and angiogenesis<sup>2</sup>



extracellular

membrane

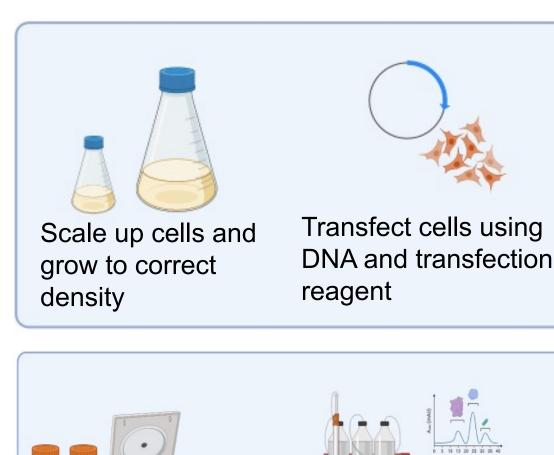
Extended closed

#### What is a minibinder?

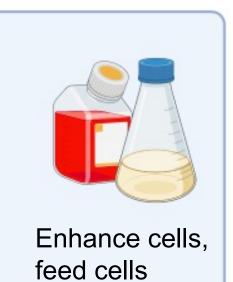
- ❖ De novo, computationally designed proteins ❖Small (~7 kDa) and stable
- protein For therapeutic application, it can be used as an alternative
- to antibody therapy Preliminary experiments have shown that V5 minibinder activates integrin



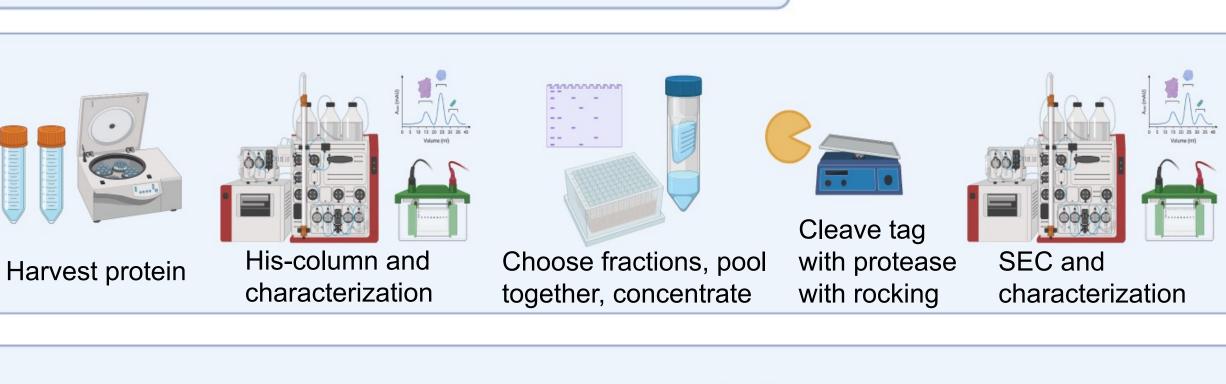
## **IETHODS**







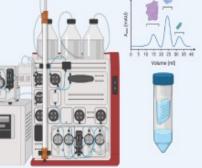
Experimental workflow for Cryogenic Electron Microscopy (Cryo-EM)



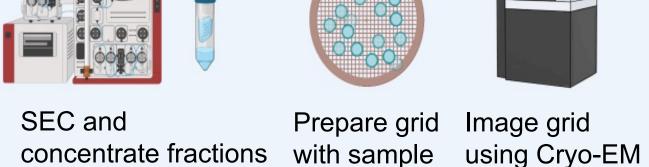


Complex protein

with minibinder





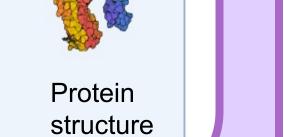








# **Process Cryo-EM**



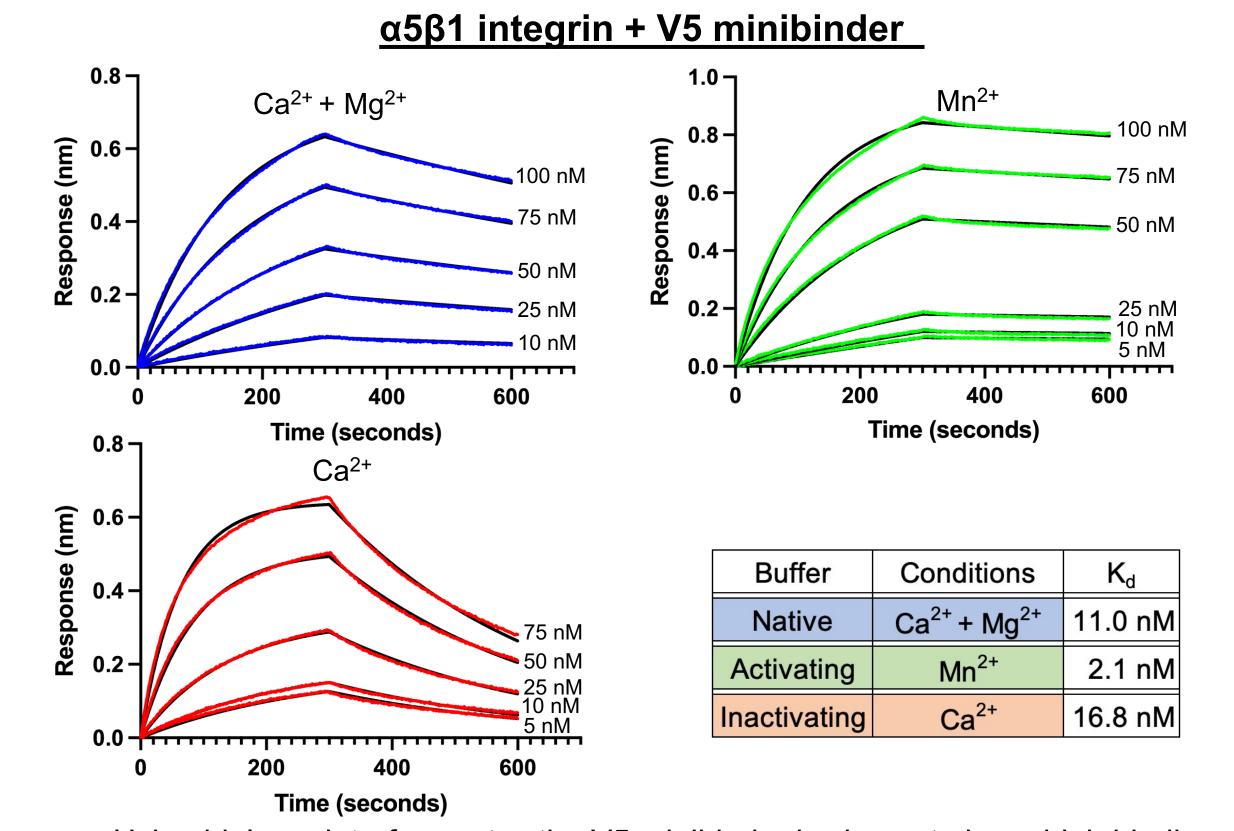
Thanks to the Campbell lab for having me, and special thanks to Rachel Werther, Jeremy Hollis, and Adam Nguyen for

Thank you to Caleigh Azumaya and rest of Cryo-EM core for helping collect Cryo-EM data and training me!

Thank you Xinru Wang and the Baker lab for collaborating and providing the minibinders!

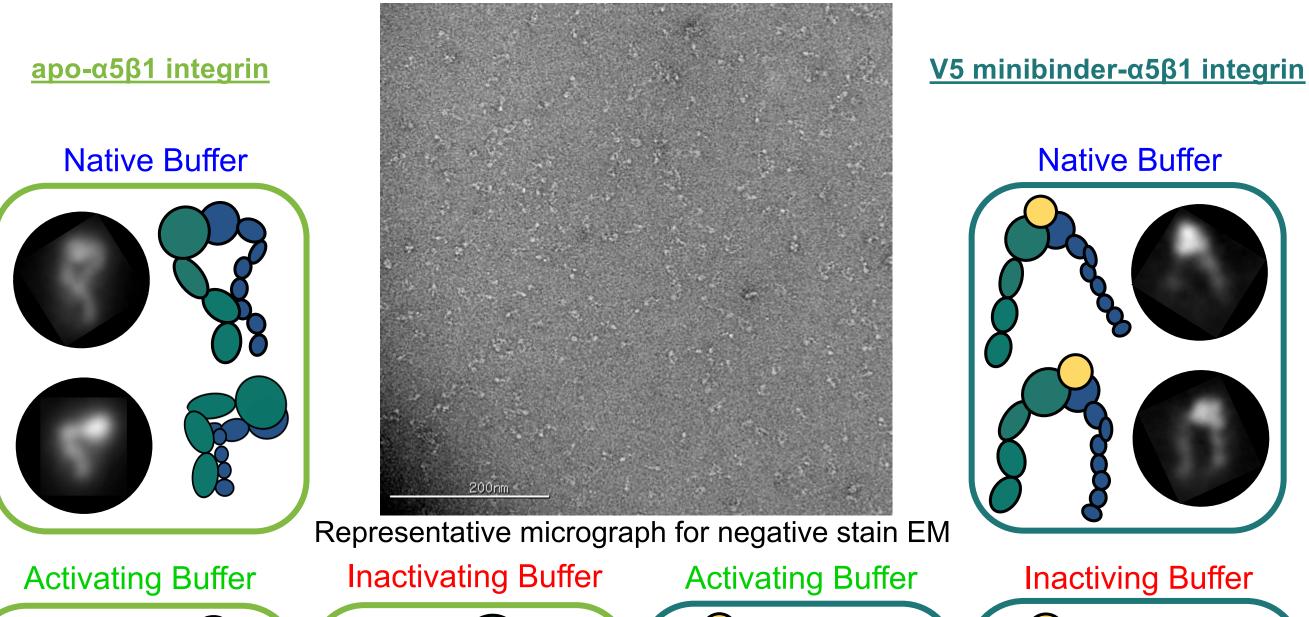
This work is funded by Fred Hutchinson Cancer Center. The Summer Undergraduate Research Program is supported in part by the Fred Hutch Internship Program and individual labs/research groups. The Campbell lab is supported by a Pew Biomedical Scholar award, the NIH MIRA award R35GM147414, and generous donations from Koss Family and Obliteride.

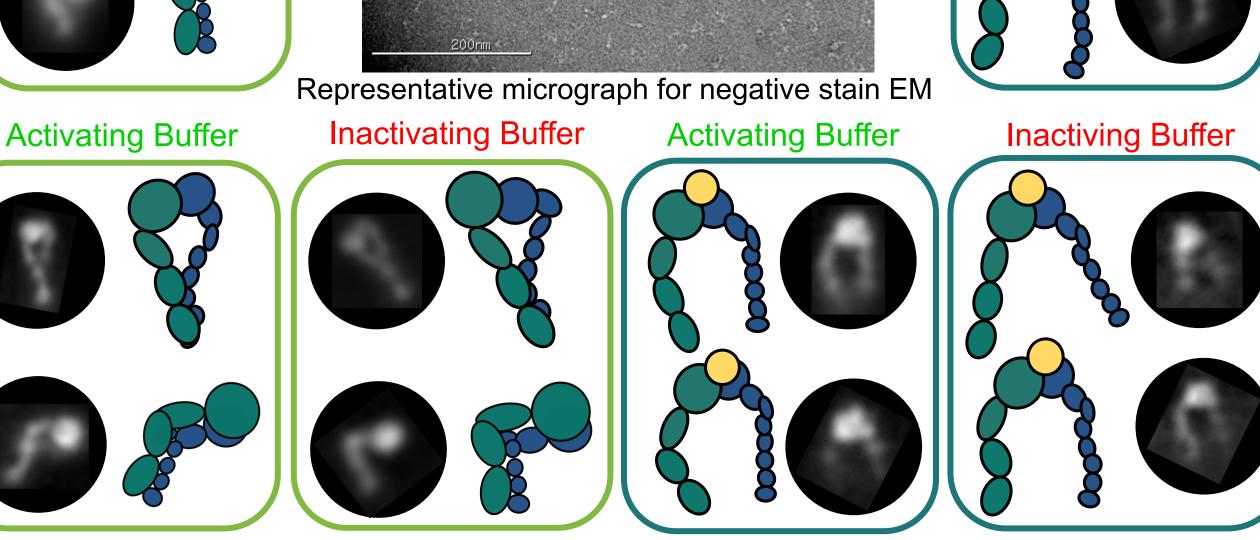
### MINIBINDER HAS HIGH BINDING AFFINITY



Using biolayer interferometry, the V5 minibinder is shown to have high binding affinity in all cation conditions and varying  $\alpha 5\beta 1$  integrin concentrations.

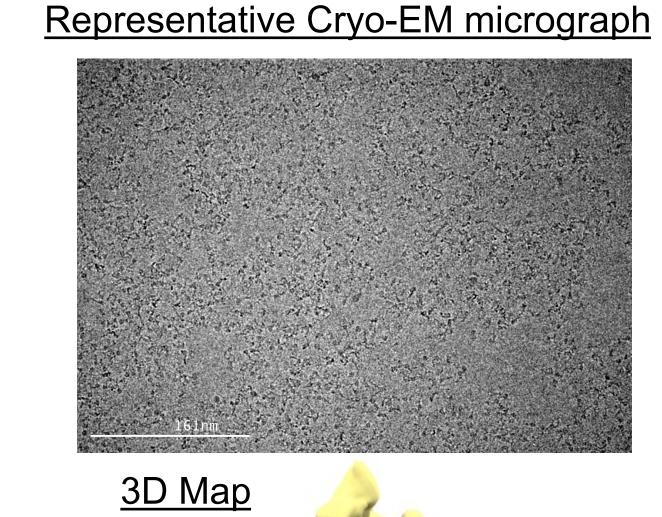
## MINIBINDER INDUCES OPEN-EXTENDED CONFORMATION

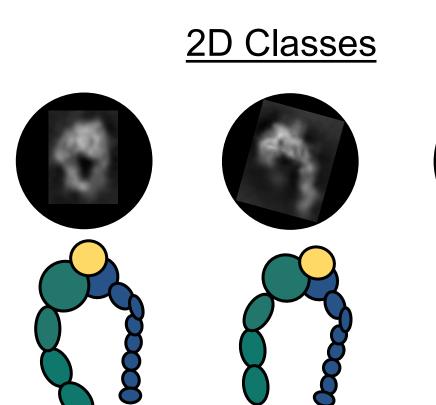


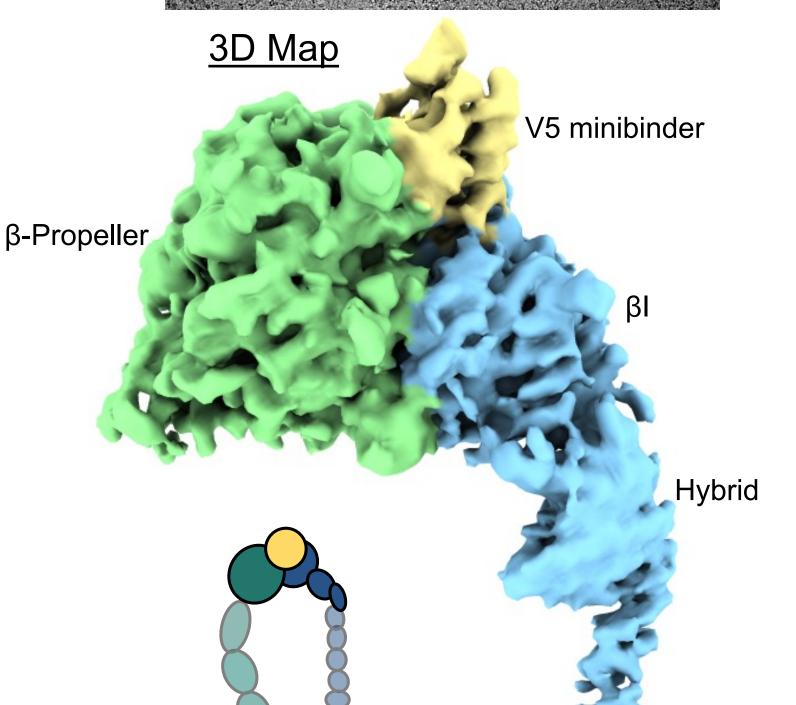


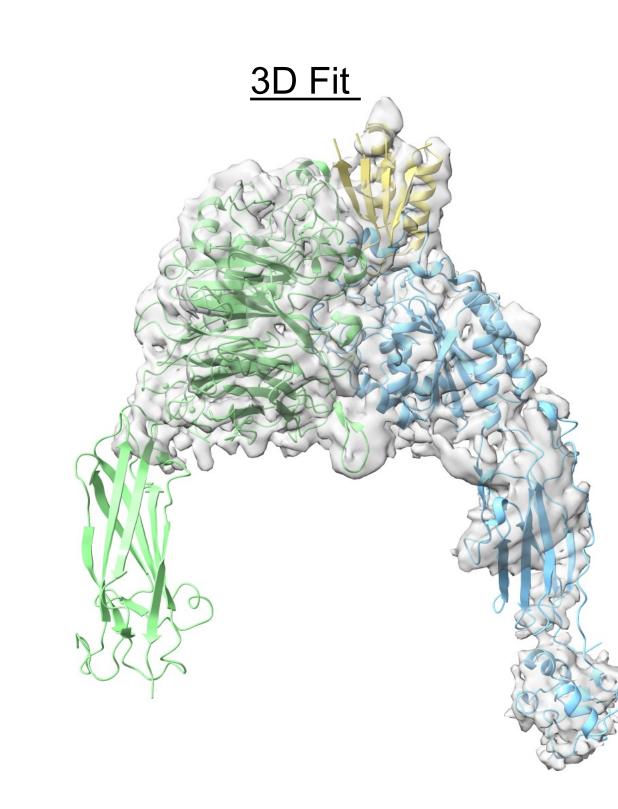
Using negative stain, conformational analysis was done on ligand-free (apo-α5β1 integrin) and ligand-bound (V5 minibinder-α5β1 integrin) α5β1 integrin under different cation conditions. Under all conditions the minibinder induced a conformational change in  $\alpha 5\beta 1$  integrin to the extended open state.

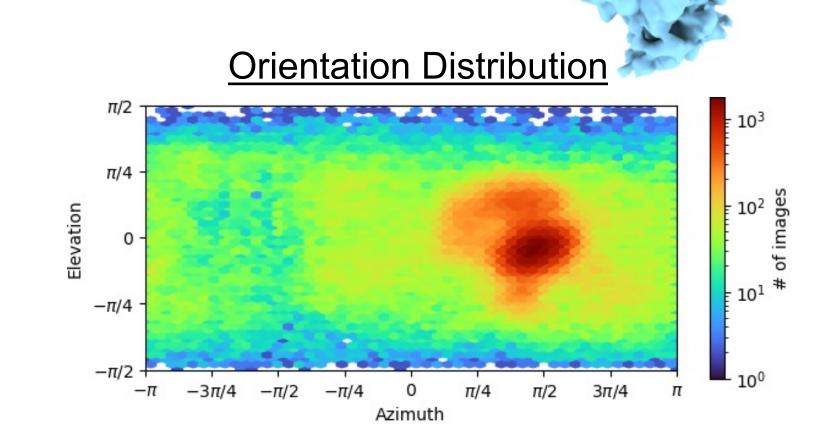
## STRUCTURAL VALIDATION USING CRYO-EM

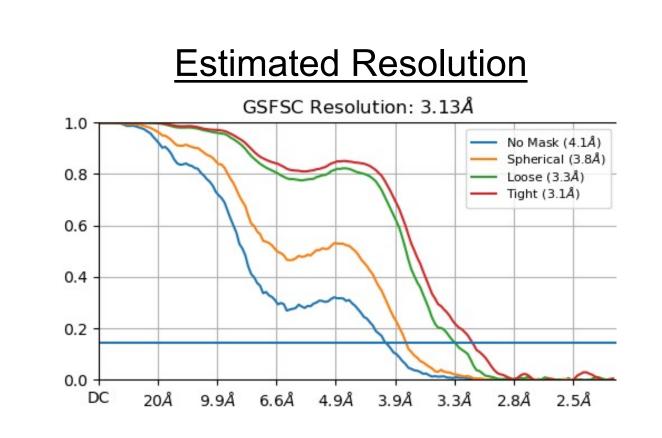








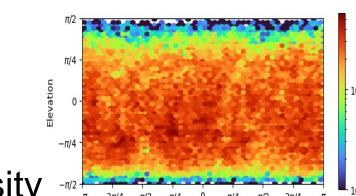




Using a Glacios electron microscope and K3 detector (camera), we verified that V5 minibinder is bound to α5β1 integrin and does induce the extended open state in inactivating buffer (Ca<sup>2+</sup>) with 3.1 Å resolution 3D map, but certain orientations are favored more (red). α5β1 integrin model from PDB ID: 7NWL.

## **FUTURE WORK**

Experiment with different sample preparation and collection methods to improve the distributions of orientations for Cryo-EM (see image on right).



- ❖Perform Cryo-EM on next-generation minibinders with improved characteristics such as higher binding affinity or specificity

## REFERENCES & ACKNOWLEDGEMENTS

1.Bachmann, M., Kukkurainen, S., Hytönen, V. P. & Wehrle-Haller, B. Cell adhesion by integrins. Physiol.

Rev. 99, 1655–1699 (2019). 2.Schumacher, S. et al. Structural insights into integrin α5β1 opening by fibronectin ligand. Sci. Adv. 7, (2021).

3. Punjani, A., Rubinstein, J. L., Fleet, D. J. & Brubaker, M. A. cryoSPARC: algorithms for rapid unsupervised cryo-EM structure determination. Nat. Methods 14, 290–296 (2017).

mentoring me, and Adam for providing apo-α5β1 data.

